



First Pharmacophoric Hypothesis for 5-HT₇ Antagonism

María L. López-Rodríguez,^{a,*} Esther Porras,^a Bellinda Benhamú,^a
José. A. Ramos,^b M. José Morcillo^c and José L. Lavandera^a

^a*Departamento de Química Orgánica I, Facultad de Ciencias Químicas, Universidad Complutense, E-28040, Madrid, Spain*

^b*Departamento de Bioquímica y Biología Molecular III, Facultad de Medicina, Universidad Complutense, E-28040, Madrid, Spain*

^c*Facultad de Ciencias, Universidad Nacional de Educación a Distancia, E-28040, Madrid, Spain*

Received 28 January 2000; accepted 13 March 2000

Abstract—In order to make the first contribution to the elucidation of essential structural features for 5-HT₇ antagonism, a set of thirty 5-HT₇ antagonists were selected from the literature. A pharmacophore model was built using Molecular Modeling studies with Catalyst program. The information contained in this model was validated with new synthesized compounds. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

Serotonin (5-hydroxytryptamine, 5-HT) was discovered over 50 years ago¹ and continues to generate interest as one of the most attractive targets for medicinal chemists. Molecular biological data have revealed the existence of fourteen serotonin receptor subtypes, which can be classified in seven families (5-HT_{1–7}).² The 5-HT₇ subtype is the most recent addition to the burgeoning family of 5-HT receptors.³ Although the biological functions of the 5-HT₇ receptor are poorly understood, preliminary evidences suggest that it may be involved in depression,⁴ control of circadian rhythms,⁵ and relaxation in a variety of vascular smooth muscles.⁶ Nevertheless, the therapeutic utility of 5-HT₇ receptor ligands awaits the development of selective agonists and antagonists. During our work only two selective 5-HT₇ receptor antagonists (SB-258719⁷ and DR4004⁸) were discovered, both from a high-throughput screening of compound libraries. In the meantime of editorial revision, Lovell et al.⁹ have reported SB-269970 as a new selective antagonist structurally related to SB-258719. Information on the structural requirements of 5-HT₇ ligands is still unknown and its determination is the major aim for developing specific compounds. In a rational drug design, identification of the pharmacophore is one of the most important steps, especially when the structure and properties of the bioreceptor

remain unknown. Therefore, our aim in this communication is to report the essential structural features for 5-HT₇ antagonism. The validation of the pharmacophore using data of new synthesized compounds suggests consistencies in structural requirements.

Pharmacophore Generation

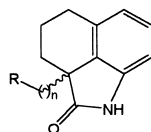
The study was performed using the software package Catalyst¹⁰ installed on a Silicon Graphics O2 workstation. A set of thirty 5-HT₇ antagonists^{7,8,11–17} structurally different from a chemical feature standpoint was selected from the reported data as the target training set for Catalyst analysis (Tables 1–4). In cases where the chirality of a stereogenic center was not specified, Catalyst generated and considered alternative stereoisomers. All structures were built de novo using 2D/3-D editor sketcher in Catalyst. Conformational models were calculated using a 15 Kcal energy cutoff (minimization convergence criteria during conformational analysis: energy convergence=0.1 Kcal/mol/Å, gradient convergence=0.01 Kcal/mol/Å). The number of conformers generated for each substrate was limited to a maximum of 250. All molecules with their associated conformations were regrouped including the biological data (pK_i). Hypothesis generation was performed and twelve hypotheses were obtained using low energy conformers of the molecules in the training set. After assessing all generated hypotheses, the most plausible one was considered the best. The goodness of the structure–activity correlation was estimated by means of r^2 .

*Corresponding author. Fax: +34-91-394-4103; e-mail: mluzlr@eucmax.sim.ucm.es

Table 1. Training set used in the generation of the 5-HT₇ antagonist pharmacophore

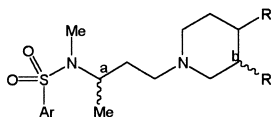
Number	Compound	p <i>K</i> _i (5-HT ₇)	References
1	Metergoline	8.2	12, 13, 14
2	Mesulergine	8.1	12
3	2-Br-LSD	8.0	11a
4	Methysergide	7.9	12
5	Clozapine	7.9	12
6	(<i>S</i>)-Methiothepin	9.0	12
7	Cyproheptadine	7.3	12
8	Mianserin	7.2	15
9	(+)-Butaclamol	7.2 ^a	11a, 15
10	Ritanserine	7.8	12
11	Spiperone	7.7	15, 16

^aThis value represents the mean of different p*K*_i values reported in refs 11a and 15.

Table 2. Training set used in the generation of the 5-HT₇ antagonist pharmacophore

Number	<i>n</i>	R	p <i>K</i> _i (5-HT ₇) ^a
12	2	4-phenylpiperazin-1-yl	7.0
13	3	4-phenylpiperazin-1-yl	8.3
14	4	4-phenylpiperazin-1-yl	8.5
15	4	4-(2-methoxyphenyl)piperazin-1-yl	8.3
16	4	4-(2-cyanophenyl)piperazin-1-yl	8.4
17	4	4-(2-pyridyl)piperazin-1-yl	8.7
18 (DR4004)	4	4-phenyl-1,2,3,6-tetrahydropyridyl	8.7
19	4	4-cyclohexylpiperazin-1-yl	5

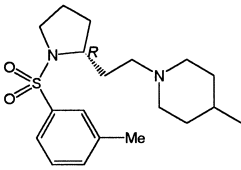
^aValues reported in ref 8.

Table 3. Training set used in the generation of the 5-HT₇ antagonist pharmacophore

Number	Ar	R	R'	Stereochemistry		p <i>K</i> _i (5-HT ₇) ^a
				a	b	
20	1-naphthyl	H	Me	<i>R,S</i>	<i>R,S</i>	7.2
21	1-naphthyl	H	Me	<i>R</i>	<i>R</i>	6.9
22	1-naphthyl	H	Me	<i>R</i>	<i>S</i>	6.2
23	1-naphthyl	H	Me	<i>S</i>	<i>R</i>	5.8
24	1-naphthyl	H	Me	<i>S</i>	<i>S</i>	5
25 (SB-258719)	3-methylphenyl	Me	H	<i>R</i>	—	7.5
26	1-naphthyl	Me	H	<i>R</i>	—	7.5
27	3,4-dichlorophenyl	Me	H	<i>R</i>	—	7.5
28	3,4-dibromophenyl	Me	H	<i>R</i>	—	7.7
29	4,5-dibromo-2-thienyl	Me	H	<i>R</i>	—	7.8

^aValues reported in ref 7.

Table 4. Training set used in the generation of the 5-HT₇ antagonist pharmacophore

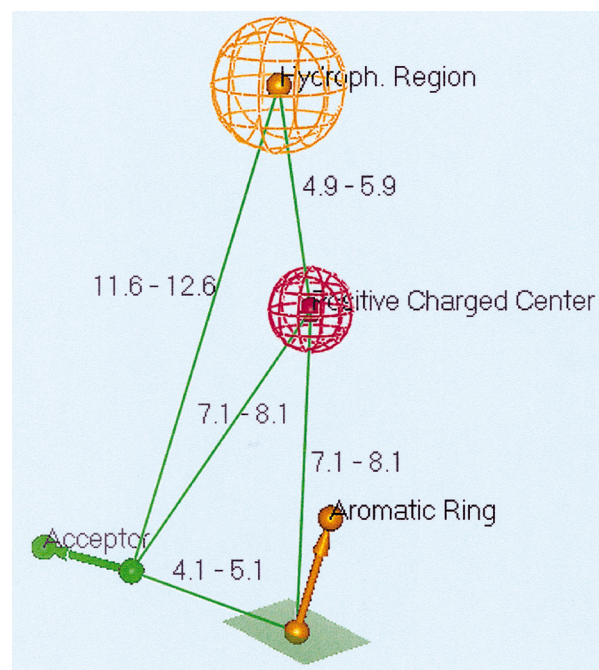
Number	Compound	p <i>K</i> _i (5-HT ₇) ^a
30		8.5

^aValue reported in ref 17.

Results

According to the hypothesis generated by catalyst, the minimal structural requirements for 5-HT₇ antagonism consist of an aromatic ring, a basic nitrogen atom (positive ionizable center), a H-bonding acceptor group and a hydrophobic region at 4.9–5.9 Å apart from the basic center (Fig. 1). For all the molecules in the training set, reasonable low-energy conformers that align on the hypothesis were found. The overall ability of this hypothesis to estimate properly the affinities of all molecules within the training set is shown by the good *r*² value between predicted and estimated affinities (*r*² = 0.921). This pharmacophoric assumption was then validated using new naphtholactam and naphthosultam derivatives (Fig. 2). Affinity data (Table 5) suggest consistencies in required structural features.

Compounds **31–45**¹⁸ were obtained by treatment of intermediates **46** with the corresponding piperazines and piperidines **47** in the presence of triethylamine and ace-

**Figure 1.** Proposed pharmacophore for 5-HT₇ antagonism.

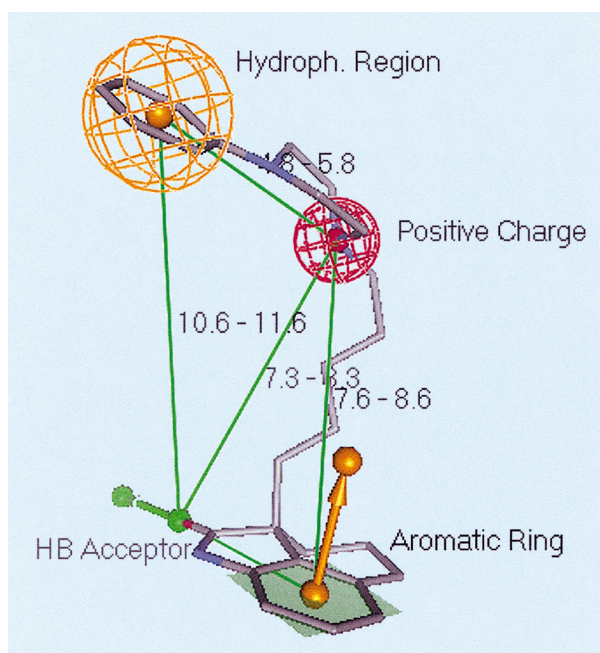
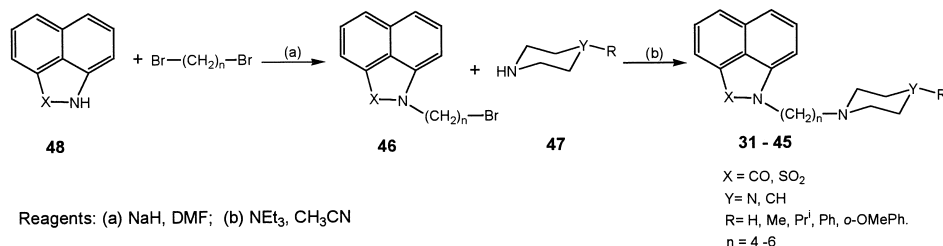


Figure 2. Compound **42** mapped on the hypothesis generated for 5-HT₇ receptor antagonists.

Table 5. Binding affinity of synthesized compounds at 5-HT₇ receptors.^a

Compd	X	n	Y	R	pK _i ^a
31	CO	4	CH	H	<5
32	CO	4	CH	Me	<6
33	CO	4	N	Me	<5
34	SO ₂	4	CH	H	<5
35	SO ₂	4	CH	Me	<5
36	SO ₂	4	N	Me	<5
37	SO ₂	5	CH	Me	<5
38	CO	4	CH	isopropyl	6.7
39	CO	4	N	<i>o</i> -methoxyphenyl	7.2
40	SO ₂	4	N	<i>o</i> -methoxyphenyl	6.7
41	CO	5	N	<i>o</i> -methoxyphenyl	7.0
42	CO	5	N	phenyl	7.1
43	SO ₂	5	N	<i>o</i> -methoxyphenyl	6.6
44	SO ₂	5	N	phenyl	6.7
45	SO ₂	6	N	<i>o</i> -methoxyphenyl	6.7

^apK_i = -log K_i. K_i (nM) values are means of two to four assays, performed in triplicate. Inhibition curves were analyzed by a computer-assisted-curve-fitting program (Prism GraphPad) and K_i values were determined from the Cheng–Prusoff equation.



Scheme 1.

tonitrile (Scheme 1). The reaction of compounds **48** with the appropriate dibromoderivative rendered key intermediates **46**. Respective hydrochloride salts of the synthesized compounds were prepared as samples for biological assays.

Target compounds were assessed for in vitro affinity at the 5-HT₇ receptor by radioligand binding assays, using [³H]-5-CT in rat hypothalamus membranes¹⁹ (Table 5). All the compounds that fit our pharmacophore model (**38–45**) show 5-HT₇ affinity (pK_i > 6.5), whereas all inactive derivatives (**31–37**, pK_i < 6.5) lack the hydrophobic moiety situated at a distance of 4.9–5.9 Å from the nitrogen atom, required for 5-HT₇ binding. These results support our pharmacophoric hypothesis. In particular, compound **42** (Fig. 2) has been selected as a new lead compound for the search for 5-HT₇ receptor ligands.

Conclusions

The pharmacophore model for 5-HT₇ antagonism described herein represents the first contribution to the rational design of agents acting at this recently identified serotonin receptor. The postulated hypothesis was validated with a series of new naphtholactam and naphthosultam derivatives that exhibit affinity for the 5-HT₇ receptor. This study offers structural insight to aid the development of novel 5-HT₇ ligands, which are essential for the knowledge of the (patho)physiological role of this serotonin receptor subtype.

Acknowledgements

This work was supported by the DGICYT (PB97-0282). The authors are grateful to U.N.E.D. for a predoctoral grant to E. Porras.

References and Notes

- Rapport, M. M.; Green, A. A.; Page, I. H. *Fed. Proc.* **1947**, *6*, 184.
- (a) In *Serotonergic Neurons and 5-HT Receptors in the CNS*; Baumgarten, H. G.; Göthert, M., Eds.; Springer-Verlag: Berlin, 1997. (b) Uphouse, L. *Neurosci. Biobehav. Rev.* **1997**, *21*, 679.
- Eglen, R. M.; Jasper, J. R.; Chang, D. J.; Martin, G. R. *Trends Pharmacol. Sci.* **1997**, *18*, 104.
- (a) Yau, J. L. W.; Noble, J.; Widdowson, J.; Seckl, J. R. *Mol. Brain Res.* **1997**, *45*, 182. (b) Shimizu, M.; Nishida, A.;

- Zensho, H.; Yamawaki, S. *J. Pharmacol. Exp. Ther.* **1996**, 279, 1551.
5. Ying, S.-W.; Rusak, B. *Brain Res.* **1997**, 755, 246.
6. (a) Terrón, J. A.; Falcon-Neri, A. *Br. J. Pharmacol.* **1999**, 127, 609. (b) De Vries, P.; De Visser, P. A.; Heiligers, J. P. C.; Villalón, C. M.; Saxena, P. R. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1999**, 359, 331.
7. Forbes, I. T.; Dabbs, S.; Duckworth, D. M.; Jennings, A. J.; King, F. D.; Lovell, P. J.; Brown, A. M.; Collin, L.; Hagan, J. J.; Middlemiss, D. N.; Riley, G. J.; Thomas, D. R.; Upton, N. *J. Med. Chem.* **1998**, 41, 655.
8. Kikuchi, C.; Nagaso, H.; Hiranuma, T.; Koyama, M. *J. Med. Chem.* **1999**, 42, 533.
9. Lovell, P. J.; Bromidge, S. M.; Dabbs, S.; Duckworth, D. M.; Forbes, I. T.; Jennings, A. J.; King, F. D.; Middlemiss, D. N.; Rahman, S. K.; Saunders, D. V.; Collin, L. L.; Hagan, J. J.; Riley, G. J.; Thomas, D. R. *J. Med. Chem.* **2000**, 43, 342.
10. catalyst 3.1; Molecular Simulations Inc.: San Diego, CA, **1996**.
11. (a) Plassat, J. L.; Amlaiky, N.; Hen, R. *Mol. Pharmacol.* **1993**, 44, 229. (b) Hirst, W. D.; Price, G. W.; Rattray, M.; Wilkin, G. P. *Br. J. Pharmacol.* **1997**, 120, 509. (c) McLean, P. G.; Coupar, I. M. *Eur. J. Pharmacol.* **1996**, 312, 215.
12. Shen, Y.; Monsma, F. J., Jr.; Metcalf, M. A.; Jose, P. A.; Hamblin, M. W.; Sibley, D. R. *J. Biol. Chem.* **1993**, 268, 18200.
13. To, Z. P.; Bonhaus, D. W.; Eglen, R. M.; Jakeman, L. B. *Br. J. Pharmacol.* **1995**, 115, 107.
14. Bard, J. A.; Zgombick, J.; Adham, N.; Vaysse, P.; Branche, T. A.; Weinshank, R. L. *J. Biol. Chem.* **1993**, 268, 23422.
15. Ruat, M.; Traiffort, E.; Leurs, R.; Tardivel-Lacombe, J.; Diaz, J.; Arrang, J.-M.; Schwartz, J.-C. *Proc. Natl. Acad. Sci. USA* **1993**, 90, 8547.
16. Jasper, J. R.; Kosaka, A.; To, Z. P.; Chang, D. J.; Eglen, R. M. *Br. J. Pharmacol.* **1997**, 122, 126.
17. Lovell, P. J., *Book of Abstracts*, XVth EFMC International Symposium on Medicinal Chemistry, Edinburgh, 6–10 September 1998, p 339.
18. New compounds were characterized by IR, ¹H, and ¹³C NMR spectroscopy and gave satisfactory combustion analysis (C, H, N). Spectral data of selected compound **42**: IR (CHCl₃, cm⁻¹) 1705 (CON), 1600, 1560, 1496, 1456 (Ar); ¹H NMR (CDCl₃) δ 1.43 (qt, *J*=7.2 Hz, 2H, CH₂), 1.58 (qt, *J*=7.5 Hz, 2H, CH₂), 1.82 (qt, *J*=7.5 Hz, 2H, CH₂), 2.35 (t, *J*=7.5 Hz, 2H, CH₂N-pip), 2.55 (t, *J*=5.1 Hz, 4H, 2CH₂-pip), 3.15 (t, *J*=5.1 Hz, 4H, 2CH₂-pip), 3.92 (t, *J*=7.5 Hz, 2H, CH₂-NCO), 6.83 (t, *J*=7.2 Hz, 1H, H₄-phenyl), 6.90 (d, *J*=7.8 Hz, 3H, H₂-, H₆-phenyl, H₈), 7.24 (t, *J*=6.9 Hz, 2H, H₃-, H₅-phenyl), 7.45 (t, *J*=8.4 Hz, 1H, H₇), 7.52 (d, *J*=8.4 Hz, 1H, H₆), 7.69 (t, *J*=8.1 Hz, 1H, H₄), 7.99 (d, *J*=8.1 Hz, 1H, H₅), 8.04 (d, *J*=7.2 Hz, 1H, H₃); ¹³C NMR (CDCl₃) δ 24.8, 26.5, 28.6 (CH₂-CH₂-CH₂), 40.1 (CH₂-NCO), 49.0 (2CH₂-pip), 53.2 (2CH₂-pip), 58.4 (CH₂N-pip), 104.8 (C₈), 115.9 (C₂-, C₆-phenyl), 119.5 (C₄-phenyl), 120.1 (C₆), 124.1 (C₃), 125.1 (C_{8b}), 126.7 (C_{2a}), 128.4, 129.0 (C₄, C₇, C₃-, C₅-phenyl), 128.6 (C_{5a}), 130.6 (C₅), 139.5 (C_{8a}), 151.2 (C₁-phenyl), 168.0 (CO); mp 211–213 °C (CH₂Cl₂:hexane)..
19. Aguirre, N.; Ballaz, S.; Lasheras, B.; Del Río, J. *Eur. J. Pharmacol.* **1998**, 346, 181.